

## Stage-specific risk stratification of diabetic retinopathy progression based on integration of OCT parameters and systemic biomarkers of inflammation and endothelial dysfunction

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## Стадія-специфічна стратифікація ризику прогресування діабетичної ретинопатії на основі інтеграції ОКТ-параметрів і системних біомаркерів запалення та ендотеліальної дисфункції

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### Abstract

**Purpose.** To assess the prognostic value of clinical and instrumental parameters and serum biomarkers of inflammation and endothelial dysfunction for the development of stage-specific risk stratification of rapid progression of diabetic retinopathy (DR) and prediction of treatment failure in patients with type 2 diabetes.

**Material and methods.** A prospective study (2 years of follow-up) included 358 patients with type 2 diabetes and DR: 189 with nonproliferative (NPDR), 96 with preproliferative

(PPDR) and 73 with proliferative (PDR). Biomarker analysis was performed in a subgroup (n=136). General clinical indicators, OCT parameters – central retinal thickness (CRT) and macular volume (MV) and serum biomarker levels – von Willebrand factor (vWF), L-selectin (LS), E-selectin (ES), endothelin-1 (ET-1), high-sensitivity C-reactive protein, endothelial-monocyte activating polypeptide II (EMAP II), endothelial NO synthase (eNOS), tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), interleukin-1 $\beta$  and -6 (IL-1 $\beta$ , IL-6), NO metabolites were determined. Patients received stage-specific treatment (conservative, laser, anti-VEGF, vitrectomy or their combination). Ineffectiveness of therapy (rapid progression of DR) was assessed using multivariate logistic regression and ROC-analysis.

**Results.** Among the clinical and instrumental parameters, the highest independent prognostic value was possessed by OCT indices, glycosylated hemoglobin (HbA1c), the presence of maculopathy and diabetic macular edema. The maximum discriminatory ability of CRT and MV was observed in PPDR (AUC 0.834 and 0.824, respectively). Among the biomarkers, the most informative were LS, EMAP II, ES, IL-1 $\beta$  and eNOS. The best stage-specific accuracy was found in: LS in NPDR (AUC 0.979), IL-1 $\beta$  in PPDR (AUC 0.990), as well as EMAP II (AUC 0.988) and eNOS (AUC 0.952) in PDR. The use of optimized thresholds allowed to identify high-risk groups, where the proportion of rapid progression exceeded 80–85% regardless of the chosen treatment method.

**Conclusion.** Treatment failure in DR in patients with T2DM is markedly stage-specific. Integration of OCT parameters with serum biomarkers of inflammation and endothelial

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dysfunction provides clinically meaningful risk stratification and can be used for early identification of patients with a high probability of rapid disease progression on standard initial therapy.

**Keywords:** diabetic retinopathy; type 2 diabetes; optical coherence tomography; biomarkers; inflammatory mediators; vascular endothelium; treatment failure.

## Резюме

**Мета.** Оцінити прогностичну цінність клініко-інструментальних параметрів і сироваткових біомаркерів запалення та ендотеліальної дисфункції для розробки стадія-специфічної стратифікації ризику швидкого прогресування діабетичної ретинопатії (ДР) і прогнозування неефективності лікування у хворих на цукровий діабет 2-го типу.

**Матеріал та методи.** У проспективне дослідження (два роки спостереження) включено 358 пацієнтів із ЦД2 та ДР: 189 із непроліферативною (НПДР), 96 із препроліферативною (ППДР) і 73 із проліферативною (ПДР). Біомаркерний аналіз проведено у підгрупі ( $n=136$ ). Визначали загальноклінічні показники, параметри ОКТ – центральну товщину (ЦТС) і макулярний об'єм (МО) та рівні сироваткових біомаркерів – фактор фон Віллебранда ( $vWF$ ), L-селектин (LS), E-селектин (ES), ендотелін-1 (ET-1), високочутливий C-реактивний протеїн, ендотеліально-моноцитарний активуючий поліпептид II (EMAP II), ендотеліальну NO-синтазу (eNOS), фактор некрозу пухлин  $\alpha$  (TNF- $\alpha$ ), інтерлейкін-1 $\beta$  та -6 (IL-1 $\beta$ , IL-

## Introduction

Diabetic retinopathy (DR) is still a major microvascular complication of diabetes mellitus (DM) and a leading cause of visual loss among working-age individuals. Despite substantial advances in screening, imaging and treatment, DR is characterized by substantial clinical heterogeneity, making prognosis in an individual patient difficult [1–4].

Current clinical guidelines provide clear recommendations for managing patients based on the stage of DR, presence of diabetic macular edema, ischemic changes and neovascularization, and using particularly optimal systemic control, intravitreal anti-vascular endothelial growth factor (VEGF) agents and vitreoretinal surgery [2, 3]. However, not all patients show an expected anatomical and clinical response even to standard treatment algorithms. This points to the limited nature of a purely phenotype-based ophthalmoscopic stratification and further necessitates a search for tools that would allow a pre-treatment separation of groups at high risk for fast disease progression and failure of initial therapeutic strategy [4, 5].

DR has been increasingly recognized not only as local microvascular injury to the retina, but also as a clinically heterogeneous neurovascular disorder involving ischemia, chronic low-grade inflammation, blood-retinal barrier

breakdown, endothelial dysfunction and microcirculation remodeling [4, 6–9]. It is for this reason that combining the two parameter groups in predictive models is of special interest. The first group of parameters reflects the local structural and functional status of the retina, primarily based on optical coherence tomography (OCT). The second group of parameters characterizes the systemic pathobiochemical processes that may model the course of retinal damage in various stages of DR [5, 6, 10].

Today, OCT is a routine tool for assessing macular changes in DR, whereas the morphometric parameters (particularly, central retinal thickness (CRT) and macular volume (MV)) and macular involvement in DR have clear clinical value for characterizing the severity of diabetic retinal lesions [6, 11, 12]. Recent reviews, however, have demonstrated that the prognostic role of particular OCT parameters is not universal, with their relationships with functional consequences and treatment response varying depending on the lesion phenotype, therapy used and method of their standardization [11, 12]. This means that even informative OCT parameters should be interpreted only in the context of the stage of DR and associated systemic mechanisms.

**Ключові слова:** діабетична ретинопатія; цукровий діабет 2-го типу; оптична когерентна томографія; біомаркери; медіатори запалення; судинний ендотелій; неефективність лікування.

Inflammation and endothelial dysfunction hold a central place among these mechanisms. Hyperglycemia, oxidative stress, accumulation of advanced glycation-end products, activation of pro-inflammatory cytokines and impaired vascular tone regulation contribute to retinal endothelial injury, increased vascular wall permeability, leukostasis, capillary occlusion and further ischemic progression [7–9]. In this context, serum biomarkers appear clinically promising, since they potentially enable minimally invasive assessment of the processes that are not always completely characterized by only ophthalmic signs [6, 10].

The most promising candidates include soluble adhesion molecules, anti-inflammatory mediators and markers of endothelial reactivity. Selectins play a key role in the early interaction of leukocytes with endothelial cells; L-selectin (LS) is associated with leukocyte activation and recruitment, whereas E-selectin (ES) reflects the activation of endothelial cells [13–15]. Soluble ES stands out as a potential predictor for development of retinopathy in type 2 diabetes [16]. Interleukin-1 $\beta$  (IL-1 $\beta$ ) is a main mediator of inflammasome-mediated inflammation that has been increasingly recognized as a component of DR progression [8, 9, 17]. Endothelial nitric oxide synthase (eNOS) is pivotal for maintaining vascular homeostasis and regulating retinal blood flow, whereas impaired nitric oxide (NO)-dependent mechanisms are considered a component of endothelial dysfunction in diabetic microvascular lesions [7, 18]. Endothelial monocyte activating polypeptide II (EMAP-II) is a multifunctional pro-inflammatory mediator that has been associated previously with microvascular diabetic complications; however, its role in risk stratification in patients with DR has been covered to a limited extent [19].

Therefore, although the data available support the concept of multimodal prediction of the course of DR, most publications are related to (a) individual biomarkers or (b) the prediction of the presence or severity of DR or the response to treatment of narrow clinical phenotypes, most commonly, diabetic macular edema (DME) [5, 6, 10–12]. However, clinically useful prediction models integrating clinical and instrumental parameters of the retina with serum inflammatory and endothelial dysfunction biomarkers and taking into account the stage of DR remain insufficiently developed.

The purpose of this study was to assess the prognostic value of clinical and instrumental markers and serum inflammatory and endothelial dysfunction biomarkers for designing the stage-specific stratification of the risk for fast progression of DR and predicting treatment failure in type 2 diabetics.

### Material and Methods

This was a single-center prospective longitudinal cohort study with a 24-month follow-up.

The procedures followed were in accordance with the ethical standards of the Helsinki Declaration (1964,

amended most recently in 2013) of the World Medical Association. The study was conducted in compliance with the requirements of the Council of Europe Convention on Human Rights and Biomedicine, and relevant laws of Ukraine. The study was approved by the Bioethics Committee of the Shupyk National Healthcare University of Ukraine.

Informed consent was obtained from all participants of the study. The inclusion criteria were 50–75-year-old male and female type 2 diabetics with any stage of DR and being born and permanently living in Ukraine. The exclusion criteria were patients with other types of diabetes; patients with severe comorbidity that was not a complication of type 2 diabetes; patients with acute or chronic dental and jaw or ear, nose and throat disorders; patients with acute or chronic infection, uveitis, congenital or acquired glaucoma, or a history of eye trauma or surgery.

Totally, 358 type 2 diabetics with DR were included in the study. Of these, a subgroup of 136 patients was used for a detailed biomarker analysis. DR was classified into nonproliferative diabetic retinopathy (NPDR; group 1; n = 189), preproliferative diabetic retinopathy (PPDR; group 2; n = 96) and proliferative diabetic retinopathy (PDR; group 3; n = 73), as per recommendations by Kohner and Porta [20, 21].

At baseline, detailed histories were taken, and clinical and demographic data (age, gender, diabetes duration expressed in years, smoking status, and systolic and diastolic blood pressures), pattern of glucose-lowering therapy used, and any systemic comorbidities (complications like polyneuropathy, diabetic foot, nephropathy, encephalopathy, microangiopathy, and/or hepatopathy) were recorded. Patients underwent eye examination including visual acuity assessment with a chart projector (CCP-3100; Huvitz Corp, Gunpo, Korea) and phoropter (HDR 7000, Huvitz Corp); static Humphrey perimetry (Humphrey Field Analyzer model 740i, Carl Zeiss Meditec Inc, Dublin, CA); refractometry with an autorefractor/ keretometer (HRK-700, Huvitz Corp); tonometry with a non-contact air-puff tonometer (HNT-7000, Huvitz Corp); corneal pachymetry with a biometer (Pentacam AXL, Oculus, Wetzlar, Germany); slit-lamp biomicroscopy (SLM-2ER, Chongqing Kanghua Ruiming Science Technology Co, Ltd, Chongqing, China); gonioscopy with a Goldmann three-mirror lens (Ocular Instruments, Bellevue, WA, USA); ophthalmoscopy with Volk digital wide-field lens (Volk Optical, Mentor, OH) and Goldmann three-mirror lens (Ocular Instruments); and OCT with RTVue RT-100 apparatus (Optovue Inc., Fremont, CA). Fundus photography (TRS-NW7SF; TOPCON, Tokyo, Japan) and fluorescein angiography were performed, if indicated. The presence of any active ocular treatment at baseline was recorded. OCT-based quantitative parameters, central retinal thickness (CRT), expressed in mm, and macular volume (MV), expressed in mm<sup>3</sup>, were determined. Manifestations of diabetic maculopathy (DMP) were assigned a binary value (ie,

“0” if absent, or “1” if present) and considered an integral characteristic of macular lesions [21, 22]. Moreover, the presence of diabetic macular edema (DME), if any, was recorded [23].

Blood samples were randomly taken in 136 patients, including 60 patients in group 1 (NPDR), 42 patients in group 2 (PPDR), and 34 patients in group 3 (PDR), to determine biomarkers. Serum levels of fasting glucose, glycosylated hemoglobin (HbA1c), total cholesterol, high-density lipoprotein cholesterol (HDL-C), low-density lipoprotein cholesterol (LDL-C), very low-density lipoprotein cholesterol (VLDL-C), triglycerides, and fibrinogen were determined colorimetrically using commercially available kits on a Cobas c311 biochemical analyzer (Roche Diagnostics GmbH, Mannheim, Germany). Coagulation hemostasis was evaluated with an extended panel of parameters including activated recalcification time (ART), prothrombin time, prothrombin index, Quick prothrombin time, international normalized ratio (INR), activated partial prothrombin time (aPPT) and thrombin time using standardized laboratory techniques [24]. Serum levels of specific markers were determined using enzyme-linked immunosorbent assays (ELISA) for advanced studies on systemic inflammation and endothelial dysfunction. Photometric measurements were performed on an ELISA plate reader (Stat Fax 303 Plus, Awareness Technology Inc, Palm City, FL). Particularly, we examined baseline levels of von Willebrand factor (vWF), LS and ES (Peninsula Laboratories, Inc., San Carlos, CA), endothelin-1 (ET-1), highly-sensitive C-reactive protein (hsCRP; Monobind Inc., Lake Forest, CA), EMAP-II (Biosource, Camarillo, CA), eNOS (BCM Diagnostics, USA), and tumor necrosis factor alpha (TNF- $\alpha$ ), IL-1 $\beta$  and IL-6 (Bender Medsystems, Vienna, Austria). Serum nitric oxide levels were determined biochemically via nitric oxide metabolism end products, nitrites (NO $_2^-$ ) and nitrates (NO $_3^-$ ), using the acidic Griess reaction [25].

In the follow-up, patients received differential initial therapy depending on the stage of the disease. Conservative treatment was administered mostly to patients in group 1 and sometimes to patients in group 2 and included hypoglycemic medications, restoration of homeostasis, and vasoprotection. Additionally, fibrates, statins and metabolic therapy were administered, if required. Groups 1 and 2 had laser treatment including panretinal laser photocoagulation (PRP) and, if required, focal laser photocoagulation (FLP). Anti-VEGF therapy was used predominantly in groups 2 and 3 and included an intravitreal anti-VEGF injection once monthly. After the first five monthly loading injections, anti-VEGF treatment intervals were determined according to a treat-and-extend regimen. Patients began receiving PRP one month after the first aflibercept injection. Up to five PRP sessions were performed, with a subsequent PRP session administered only in the presence of ischemic areas. Laser settings utilized pulse durations from 100 to 200 ms, pulse-to-pulse

periods of 100 to 150 ms, and powers from 100 to 200 mW.

Surgical treatment was performed mostly in patients in group 3 and included a 25-G three-port subtotal pars plana vitrectomy (PPV) plus PRP session, epiretinal membrane removal and endotamponade with 18% C3F8 or silicone oil 5700 centistokes, depending on the stage of the process.

A combination of intravitreal anti-VEGF medications, PRP and 25-G subtotal PPV was used mostly in patients in groups 2 and 3.

No DR progression (grade 0) was defined as the stability of ophthalmological characteristics at two years. Slow DR progression (grade 1) was defined as the worsening of some ophthalmological characteristics at two years. Fast DR progression (grade 2) was defined as worsening from NPDR to PPDR, worsening from PPDR to PDR, or the substantial worsening of the most ophthalmological characteristics in patients with PDR at two years. Treatment effect was defined as no DR progression or slow DR progression at 2 years. Treatment failure was defined as fast DR progression at 2 years.

Statistical analysis was performed with EZR version 1.54 (Saitama Medical Center, Jichi Medical University, Saitama, Japan), which is a graphical interface for R (The R Foundation for Statistical Computing, version 4.03, R Foundation for Statistical Computing, Vienna, Austria) [26]. A complete case analysis was used to address missing data. Significance was established at  $P < 0.05$  for all two-tailed tests. The analysis included three stages and was aimed at (1) stratifying patients with each DR stage into progression risk and (2) comparing various treatment strategies for efficacy in the selected subgroups. In phase 1 of the statistical analysis, binary logistic regression was used to assess the prognostic value of each marker with regard to fast DR progression (grade 2 versus grades 0 to 1), with correction for age, gender, diabetes duration, DR stage at baseline, and the presence of active initial treatment. The informativeness of the marker was assessed by an increase in the likelihood ratio (LR)  $\chi^2$ , compared to the basic model, with the Benjamini-Hochberg false discovery rate (FDR) applied for multiple comparisons. In phase 2 of the statistical analysis, stage-specific Receiver Operating Characteristic (ROC) analysis was conducted for the most informative markers: ROC curves were built separately for NPDR, PPDR and PDR, which allowed removing the effect of stage-to-stage variability in the distribution of markers. For each subgroup, the area under curve (AUC) and 95% confidence interval (CI) were calculated by the DeLong method, and the Youden's index was used to determine cut-off values. In phase 3 of the statistical analysis, patients with each stage of DR were divided into high-risk and low-risk subgroups based on the stage-specific marker threshold value. The percentage of patients with fast DR progression was calculated in each of the subgroups created for each of the methods of initial treatment. Data were presented as descriptive portions;

the difference between methods as a result of comparison was not interpreted as evidence of superiority of one of the methods due to the absence of randomization.

## Results

During prospective observation of a complete cohort ( $n = 358$ ) and the subgroup selected for biomarker analysis ( $n = 136$ ), we assessed the clinical trajectory of DR after the initial treatment was administered. In phase 1 of our statistical analysis, in order to identify the most significant predictors of treatment failure (fast DR progression), we used a binary-regression-based screening analysis, with correction for age, gender, diabetes duration, DR stage at baseline, and the presence of active primary treatment. A hierarchy of informativeness of the parameters examined was established based on the increase in the LR  $\chi^2$ , with the FDR applied for multiple comparisons.

OCT parameters, CRT and MV, were found to have the highest prognostic value among general clinical and ophthalmological markers, which indicated the development of various manifestations of DMP ( $q < 0.05$  for all comparisons). Characteristics of endothelial dysfunction and inflammation (LS, EMAP II, ES, IL-1 $\beta$  and eNOS) showed the highest prognostic potential among the biomarkers determined. It is these characteristics that were selected for further, more detailed analysis.

In phase 2 of our statistical analysis, stage-specific ROC analysis was conducted to quantitatively and accurately assess the prognostic value of the selected markers and determine cut-off points (Table 1).

Prognostic accuracy of general clinical parameters substantially depended on the stage of DR at baseline (Table 1). Thus, the discriminative capacity of the quantitative characteristics of macular edema (CRT and MV) was high in PPDR (AUC > 0.82) but moderate in NPDR and PDR.

A similar stage-specific pattern was observed in the analysis of biomarkers (Table 2).

Serum biomarkers demonstrated higher prognostic accuracy (with AUC > 0.82 for most of the biomarkers; Table 2) compared to general clinical characteristics. Interestingly that, among the serum biomarkers, L-selectin had the highest accuracy in NPDR, whereas EMAP II and IL-1 $\beta$  had the highest accuracy (AUC > 0.94) in PPDR and PDR. The Youden's index-based cut-off values for each marker in each DR stage made the basis for the final phase of the study, stage-specific stratification of the risk for fast progression of DR.

In phase 3 of our statistical analysis, stage-specific thresholds were used to divide the cohort into low-risk and high-risk subgroups, and the efficacy of various initial treatment methods was assessed. Figure 1 shows the examples of implementing this approach for the best instrumental marker (CRT) and the best early biomarker (L-selectin).

The analysis of stratified results found two stable and clinically significant patterns that were reproduced

regardless of whether we used an instrumental marker or a biomarker for discrimination. First, in high-risk subgroups (at any stage of DR), the percentage of patients with fast DR progression (i.e., treatment failure) was critically high and predominantly exceeded 80–85%. This pattern was observed regardless of the type of treatment strategy selected (conservative, laser, surgical, anti-VEGF therapy or combined), indicating an aggressive phenotype of the disease which was resistant to standard treatments. Second, a completely different pattern was seen in low-risk subgroups, with substantially better treatment response. Additionally, in low-risk subgroups of patients with PPDR and PDR, surgical or combined treatment strategy was associated with a smaller percentage of patients with fast DR progression compared to monotherapy strategy. Therefore, the use of determined stage-specific cut-off values enabled clearly identifying a cohort of patients who are highly likely to fail conventional treatment algorithms, which may justify the need for revising these algorithms or adjusting them early.

## Discussion

This study demonstrated that the prediction of initial treatment failure (which corresponded to fast DR progression in this study) in type 2 diabetics with DR cannot be based on a set of features universally applicable to all stages of DR. The major finding was that of a stage-dependent change in the most prognostically informative parameters. OCT parameters were found to be the most prognostically informative among clinical and instrumental parameters; among serum biomarkers, LS showed the best discrimination in patients with NPDR, whereas EMAP-II, IL-1 $\beta$ , ES and eNOS, in patients with PPDR and PDR. This agrees well with a current concept of DR as a dynamic neurovascular condition in which local retinal structural changes are combined with systemic signs of inflammation and endothelial dysfunction while the prognostic weight of individual markers changes with disease progression [5, 6, 10, 27, 28].

It seems pathophysiologically intuitive that we found that CRT, MV and other clinical signs of DMP had the highest independent values among general clinical and ophthalmological characteristics in our cohort of patients. The superiority of OCT parameters over HbA1c indicates that direct signs of retinal lesion activity are more important than average metabolic control characteristics in the prognosis of treatment failure. It is especially telling that CRT and MV showed the best discriminative capacity in patients with PPDR. It is likely that it is at this stage of DR macular edematous and ischemic components are sufficiently marked to exert an effect on further clinical trajectory, but are not yet “overlapped” by a complex array of fibrous proliferative and tractional changes characteristic of the later stage, PDR. Such an interpretation is consistent with understanding a phenotype-dependent role of OCT parameters in diabetic macular lesions [11, 12, 29, 30]. On the other hand, the development of these unfavorable

**Table 1.** Stage-specific results of ROC analysis of general clinical markers for predicting fast progression of diabetic retinopathy (a complete cohort, n = 358)

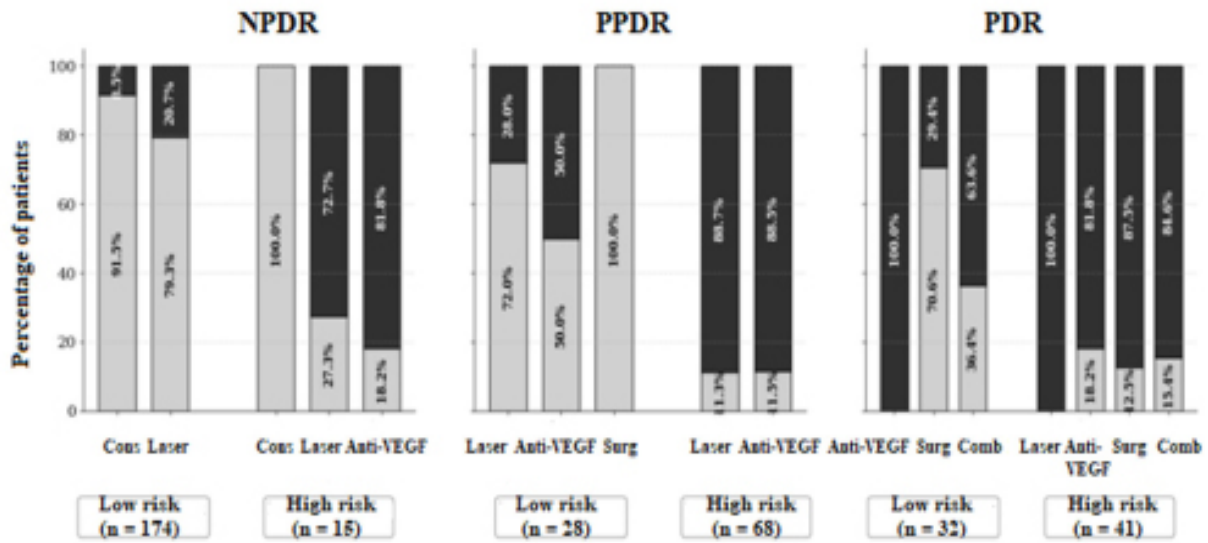
Marker	Stage of DR	n	AUC (95% CI)*	Optimal cut-off	Sensitivity, %	Specificity, %	Youden J
MV, mm <sup>3</sup>	NPDR	189	0.683 (0.576–0.789)	> 0.23	31.2	96.8	0.281
	PPDR	96	0.824 (0.733–0.915)	> 0.21	89.4	70.0	0.594
	PDR	73	0.664 (0.518–0.811)	> 0.35	60.8	77.3	0.381
CRT, $\mu$ m	NPDR	189	0.682 (0.576–0.788)	> 298	31.2	96.8	0.281
	PPDR	96	0.834 (0.747–0.921)	> 271	89.4	70.0	0.594
	PDR	73	0.659 (0.507–0.810)	> 317	68.6	72.7	0.414
DMP	NPDR	189	0.634 (0.551–0.717)	= 1	31.2	95.5	0.268
	PPDR	95	0.705 (0.606–0.804)	= 1	67.7	73.3	0.410
	PDR	73	0.707 (0.592–0.822)	= 1	68.6	72.7	0.414
DME	NPDR	189	0.615 (0.538–0.692)	= 1	25.0	98.1	0.231
	PPDR	96	0.529 (0.439–0.619)	= 1	25.8	80.0	0.058
	PDR	73	0.560 (0.434–0.686)	= 1	52.9	59.1	0.120
HbA1c, %	NPDR	189	0.604 (0.497–0.711)	> 7.7	78.1	42.7	0.208
	PPDR	96	0.693 (0.587–0.800)	> 9.6	62.1	80.0	0.421
	PDR	73	0.574 (0.434–0.714)	> 11.2	56.9	63.6	0.205

Notes: From the ROC analysis, optimal cut-off values were calculated using the Youden Index. AUC 95% confidence intervals were calculated using the DeLong method. Qualitative characteristics were represented with binary variables, such as maculopathy and DME, with “= 1” denoting the presence of a feature. Abbreviations: AUC, area under curve; CI, confidence interval; CRT, central retinal thickness; DME, diabetic macular edema; DMP, diabetic maculopathy; DR, diabetic retinopathy; NPDR, non-proliferative diabetic retinopathy; PDR, proliferative diabetic retinopathy; PPDR, pre-proliferative diabetic retinopathy; ROC, receiver operating curve

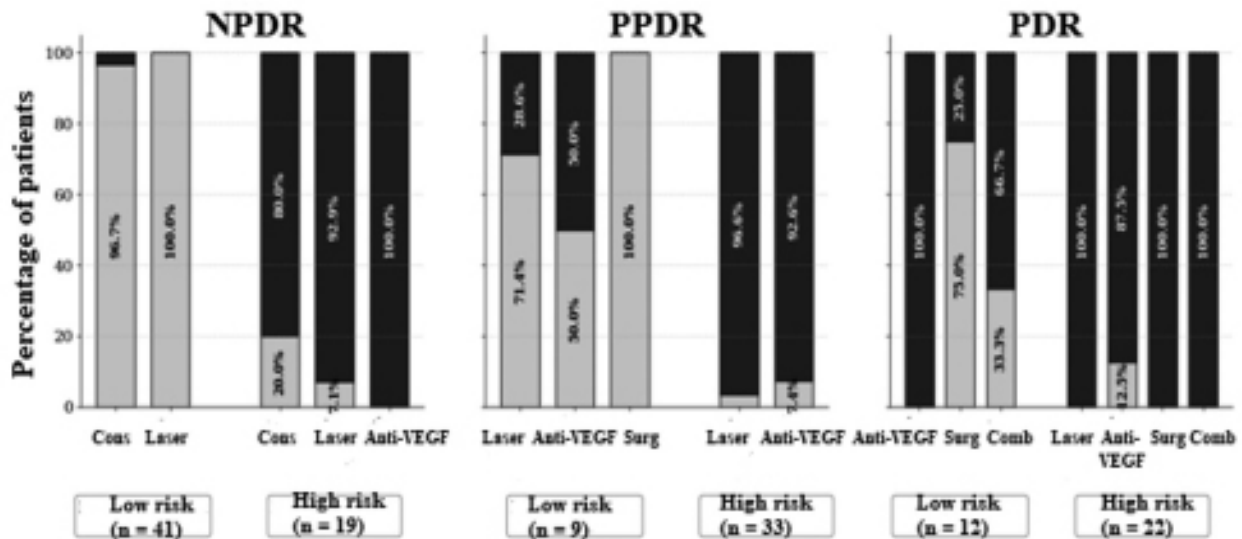
**Table 2.** Stage-specific results of ROC analysis of biomarkers of inflammation and epithelial dysfunction for predicting fast progression of diabetic retinopathy (a biomarker subgroup, n = 136)

Marker	Stage of DR	n	AUC (95% CI)*	Optimal cut-off	Sensitivity, %	Specificity, %	Youden J
LS, ng/mL	NPDR	60	0.979 (0.949–1.000)	> 33.4	94.4	95.2	0.897
	PPDR	42	0.929 (0.846–1.000)	> 49.0	93.9	77.8	0.717
	PDR	34	0.774 (0.551–0.997)	> 59.6	75.0	83.3	0.583
EMAP II, ng/mL	NPDR	60	0.769 (0.648–0.889)	> 3.59	94.4	54.8	0.492
	PPDR	42	0.943 (0.878–1.000)	> 7.04	84.8	100.0	0.848
	PDR	34	0.988 (0.960–1.000)	> 8.65	92.9	100.0	0.929
ES, ng/mL	NPDR	60	0.695 (0.514–0.876)	> 32.5	61.1	88.1	0.492
	PPDR	42	0.902 (0.807–0.997)	> 50.0	84.8	100.0	0.848
	PDR	34	0.920 (0.820–1.000)	> 58.6	89.3	83.3	0.726
IL-1 $\beta$ , pg/mL	NPDR	60	0.580 (0.403–0.757)	> 64.5	33.3	92.9	0.262
	PPDR	42	0.990 (0.968–1.000)	> 77.2	97.0	100.0	0.970
	PDR	34	0.917 (0.822–1.000)	> 192.2	82.1	100.0	0.821
eNOS, pmol/mL	NPDR	60	0.708 (0.566–0.851)	< 206	72.2	73.8	0.460
	PPDR	42	0.747 (0.599–0.896)	< 193	57.6	100.0	0.576
	PDR	34	0.952 (0.883–1.000)	< 121	89.3	100.0	0.893

Notes: From the ROC analysis, optimal cut-off values were calculated using the Youden Index. \*AUC 95% confidence intervals were calculated using the DeLong method. Abbreviations: AUC, area under curve; CI, confidence interval; DR, diabetic retinopathy; EMAP II, endothelial-monocyte-activating polypeptide II; eNOS, endothelial nitric oxide synthase; ET-1, endothelin-1; IL-1 $\beta$  interleukin-1 $\beta$ ; NPDR, non-proliferative diabetic retinopathy; PDR, proliferative diabetic retinopathy; PPDR, pre-proliferative diabetic retinopathy; ROC, receiver operating curve



**Fig. 1.** Efficacy of methods of initial treatment for diabetic retinopathy depending on stage-specific stratification of the risk of diabetic retinopathy progression based on central retinal thickness (CRT). Light-grey bar segments represent percentages of patients with no or slow disease progression (grades 0 to 1), and dark-grey bar segments, percentages of those with fast disease progression (grade 2; treatment failure) in a complete cohort (n = 358). Separation into low-risk and high-risk subgroups was based on stage-specific CRT cut-off values determined by ROC analysis:  $\leq 298$  and  $> 298$   $\mu\text{m}$  for NPDR;  $\leq 271$  and  $> 271$   $\mu\text{m}$  for PPDR; and  $\leq 317$  and  $> 317$   $\mu\text{m}$  for PDR, respectively. The n values under the abscissa represent numbers of patients in each risk subgroup. Comparisons of treatment strategies are only descriptive in nature. Abbreviations: Cons, conservative treatment; Comb, combination treatment; DR, diabetic retinopathy; NPDR, non-proliferative diabetic retinopathy; PDR, proliferative diabetic retinopathy; PPDR, pre-proliferative diabetic retinopathy; Surg, surgical treatment.



**Fig. 2.** Efficacy of methods of initial treatment for diabetic retinopathy depending on stage-specific stratification of the risk of diabetic retinopathy progression based on serum L-selectin (LS) level. Light-grey bar segments represent percentages of patients with no or slow disease progression (grades 0 to 1), and dark-grey bar segments, percentages of those with fast disease progression (grade 2; treatment failure) in the biomarker subgroup (n = 136). Separation into low-risk and high-risk subgroups was based on stage-specific LS cut-off values determined by ROC analysis:  $\leq 33.4$  and  $> 33.4$  ng/mL for NPDR;  $\leq 49.0$  and  $> 49.0$  ng/mL for PPDR;  $\leq 59.6$  and  $> 59.6$  ng/mL for PDR, respectively. The n values under the abscissa represent numbers of patients in each risk subgroup. Comparisons of treatment strategies are only descriptive in nature. Abbreviations: Cons, conservative treatment; Comb, combination treatment; DR, diabetic retinopathy; NPDR, non-proliferative diabetic retinopathy; PDR, proliferative diabetic retinopathy; PPDR, pre-proliferative diabetic retinopathy; Surg, surgical treatment.

changes in the retinal structure of patients with DR is closely associated with systemic metabolic abnormalities. As we have demonstrated in our recent study, atherogenic dyslipidemia (particularly, increased LDL-C, VLDL-C and non-HDL-C levels) is a “profound” systemic predictor of the development of DME and worsened OCT parameters [31]. This further underlines the need for a comprehensive approach to risk stratification, with the involvement of local retinal characteristics and the systemic metabolic profile.

We found LS to be the most informative serous marker of early risk stratification in DR. This observation has a convincing biological basis since (1) selectins are key mediators in the early interaction of leukocytes with endothelial cells, and (2) leukostasis is believed to be an early component of retinal microvascular damage in diabetes [7, 13–15, 28, 32–35]. A high prognostic value of LS in NPDR may reflect the situation when irreversible proliferative changes are not yet clinically dominant, but mechanisms of leukocyte adhesion, local inflammation and blood-retinal barrier breakdown are already actively involved in the disease process. While demonstrating the role of HbA1c, lipoprotein cholesterol and coagulation characteristics in DR progression, we have previously pointed out that it is at the non-proliferative stage of DR that identifying predictors of this progression is especially important [36]. Findings of the current study complement these results, demonstrating that, among the specialized panel of inflammation and endothelial dysfunction markers, it is soluble L-selectin that is the earliest and most sensible indicator of the transition to an unfavorable course of DR. An increased role of ES in PPDR and PDR is also reasonable because this marker directly reflects the activation of endothelial cells proper, that is, the vascular stress that expectedly increases with increasing severity of DR [7, 13–15, 28, 32–35].

Our results, however, should not be interpreted as the confirmation of the absolute universality of circulating adhesion molecules in all populations of patients. Some studies found no persuasive association between soluble adhesion molecules and the development of DR [16]. Most likely, such variability may be due to differences in design, type of diabetes, distribution of DR stages in the sample, selected endpoints and/or no division of the sample into groups with various baseline stages of DR. It is telling that prospective and cohort studies, however, support the prognostic value of endothelial dysfunction and low-grade inflammation in the development and progression of DR [16, 28, 37–39]. In this context, our results do not contradict previous data, but rather clarify the conditions under which these markers are actually clinically informative.

More important appears to be a shift in “leading biomarkers” with the transition from NPDR to PPDR and PDR. High discriminative capacity of EMAP-II and IL-1 $\beta$  in these stages of DR supposes that, with the progression of DR, a chronic pro-inflammatory environment and a deeper endothelial lesion become more

decisive than early leukocyte adhesion. This interpretation is especially convincing for IL-1 $\beta$ , because this cytokine is a central effector of the inflammasome-mediated response associated with pyroptosis, increased vascular permeability and retinal lesion progression [8, 9, 17, 40]. Although data are still limited on the role of EMAP-II in the context of DR, it is known to mediate the interaction of monocytes with endothelial cells and be involved in apoptosis and pathological vascular remodeling [19, 41, 42]. This is why its high discriminative capacity in PPDR and PDR is one of the most interesting findings of this study. Equally telling was the finding that low eNOS was an informative predictor of PDR. This is consistent with the idea of the depletion of NO-dependent vasoprotective mechanisms, loss of endothelial reactivity and loss of vascular homeostasis under conditions of severe microangiopathy [7, 18]. The present study is a logical continuation of our previous study [43] in which we (1) developed a model for predicting the efficacy of treating DR in type 2 diabetes on the basis of determination of markers of endothelial dysfunction and (2) demonstrated high sensitivity and specificity of EMAP II and eNOS. The present study, however, substantially expands this understanding by demonstrating that the prognostic values of these endothelial factors are not constant and it is in late stages (pre-proliferative and proliferative) of DR that they reach their maximum.

Taken together, our findings allow us to build a consistent pathobiological scheme, from early leukocyte-endothelial activation (to which LS is sensitive) via an increased endothelial response with an increased expression of ES, to persistent inflammatory amplification associated with IL-1 $\beta$  and EMAP-II, and, finally, to the depleted endothelial reserve which is reflected by a reduced expression of eNOS.

This scheme is backed by experimental evidence: leukostasis, intercellular adhesion and inflammation are directly involved in the development of vascular permeability, capillary occlusion and endothelial cell death in diabetic retina [34, 44–46]. The practical implications of our findings are not so much the selection of the best method of treatment of DR as the identification of the aggressive phenotype of DR prior to therapy initiation. The fact that the percentage of patients with fast DR progression in high-risk subgroups predominantly exceeded 80–85% regardless of the type of initial treatment strategy selected indicates the limitedness of a conventional step-by-step approach to treatment strategy selection in this category of patients. At the same time, better outcomes in low-risk than high-risk subgroups and lower frequency of fast DR progression in patients with PPDR/PDR that received surgical or combined treatment strategy should be interpreted carefully, because, in this study, comparisons of treatment strategies were only descriptive in nature. However, the very possibility of separating patients for whom early revision of treatment strategy, more intensive

monitoring or multimodal management may be appropriate would have a direct clinical value [2, 3, 5, 9, 11, 30].

Advantages of this study include its prospective design, a clinically relevant endpoint, combination of general clinical data with a detailed biomarker analysis in a biomarker subgroup, and determination of practical cut-off values for each DR stage.

Results of the present study should be interpreted with some limitations. First, because the biomarker subgroup was of a moderate size, further studies with larger sample sizes are required to confirm the accuracy of individual assessments. Second, the levels of all the serum markers were determined at baseline only, whereas the dynamics of these levels in the presence of treatment would add some prognostic value. Third, the proposed cut-off values are of a pragmatic nature for this cohort and should be externally validated prior to wide implementation. Finally, because this work did not aim to compare individual treatment strategies for efficacy, any conclusions with regard to advantages of a particular intervention would be incorrect. The present study, however, demonstrates that integration of OCT parameters with serum biomarkers of inflammatory and endothelial dysfunction may provide a basis for more accurate personalized stratification of the risk of initial treatment failure in DR.

### Conclusions

First, in type 2 diabetics with DR, the prognostic value of markers of initial treatment failure (which in this study was defined as fast disease progression) is of an apparently stage-specific nature; using a universal set of predictors for all stages of DR will be insufficiently informative.

Second, among clinical ophthalmological parameters, OCT parameters (CRT and MV) and signs of various manifestations of DMP were found to have the highest independent prognostic value, with CRT and MV demonstrating the highest discriminative capacity in the PPDR stage of DR.

Third, serum biomarkers of inflammatory and endothelial dysfunction generally demonstrated a higher prognostic accuracy than general clinical characteristics. LS was the most informative marker in NPDR; IL-1 $\beta$  and EMAP II, in PPDR, whereas EMAP II and eNOS, in PDR.

Fourth, the determination of the stage-specific cut-off values for the most informative clinical-and-instrumental and laboratory markers provided a separation of patients into subgroups with a low and high risk of initial treatment failure, respectively, with the percentage of patients with fast DR progression in high-risk subgroups predominantly exceeding 80–85%.

Finally, integrating OCT parameters with serum biomarkers of inflammatory and endothelial dysfunction provides a justified approach to personalized stage-specific stratification and may be used for early separation of DR patients with a high probability of initial treatment failure.

### Author contributions

SYM: Conceptualization, Project Administration, Data Analysis and Interpretation; AVS: Data Curation, Investigation, Data Analysis and Interpretation, Writing - original draft preparation, Writing - Review and Editing; VNS: Data Curation, Investigation; BMS: Investigation, Data Analysis and Interpretation, Writing - original draft preparation; SVZ: Conceptualization, Data Analysis and Interpretation. All authors have read and approved the final version of the manuscript.

### Ethical Declaration

This study involved human subjects, was approved by the local bioethics committee, and followed ethical standards as outlined in the Declaration of Helsinki. Informed consent was obtained from all subjects. This study did not include animal experiments.

### Disclaimer

This manuscript reflects the views of the authors and may not reflect the views of their institutions.

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None.

### Conflict of interest

All authors declare no conflict of interest that could influence their views on the subject matter or materials described and discussed in this manuscript.

### Data Availability Declaration

All the data obtained or examined during this study has been incorporated into this published article.

### Abbreviations

AUC, area under curve; CI, confidence interval; DME, diabetic macular edema; DM, diabetic mellitus; DR, diabetic retinopathy; EMAP II, endothelial-monocyte-activating polypeptide II; eNOS, endothelial nitric oxide synthase; ET-1, endothelin-1; HbA1c, glycated hemoglobin; hs-CRP, highly sensitive C-reactive protein; IL-1 $\beta$ , interleukin 1 $\beta$ ; IL-6, interleukin 6; NO, nitric oxide; NOx, nitric oxide metabolites; NPDR, non-proliferative diabetic retinopathy; OCT, optical coherence tomography; OR, odds ratio; PDR, proliferative diabetic retinopathy; PPDR, pre-proliferative diabetic retinopathy; PRP, panretinal laser photocoagulation.

### References

1. Teo ZL, Tham YC, Yu M, Chee ML, Rim TH, Cheung N, et al. Global Prevalence of Diabetic Retinopathy and Projection of Burden through 2045: Systematic Review and Meta-analysis. *Ophthalmology*. 2021 Nov;128(11):1580-1591. doi: 10.1016/j.ophtha.2021.04.027.
2. Lim JI, Kim SJ, Bailey ST, Kovach JL, Vemulakonda GA, Ying GS, Flaxel CJ; American Academy of Ophthalmology Preferred Practice Pattern Retina/Vitreous Committee. Diabetic Retinopathy Preferred Practice Pattern®. *Ophthalmology*. 2025 Apr;132(4):P75-P162. doi: 10.1016/j.ophtha.2024.12.020.
3. American Diabetes Association Professional Practice Committee for Diabetes\*. 12. Retinopathy, Neuropathy, and Foot Care: Standards of Care in Diabetes-2026. *Diabetes Care*.

- 2026 Jan 1;49(Supplement\_1):S261-S276. doi: 10.2337/dc26-S012.
4. Kaurich C, Mahajan N, Bhatwadekar AD. Precision Medicine for Diabetic Retinopathy: Integrating Genetics, Biomarkers, Lifestyle, and AI. *Genes (Basel)*. 2025 Sep 16;16(9):1096. doi: 10.3390/genes16091096.
  5. Mellor J, Jeyam A, Beulens JWJ, Bhandari S, Broadhead G, Chew E, et al. Role of Systemic Factors in Improving the Prognosis of Diabetic Retinal Disease and Predicting Response to Diabetic Retinopathy Treatment. *Ophthalmol Sci*. 2024 Feb 17;4(4):100494. doi: 10.1016/j.xops.2024.100494.
  6. Chondrozoumakis G, Chatzimichail E, Habra O, Vounotrypdis E, Papanas N, Gatziofufas Z, et al. Retinal Biomarkers in Diabetic Retinopathy: From Early Detection to Personalized Treatment. *J Clin Med*. 2025 Feb 18;14(4):1343. doi: 10.3390/jcm14041343.
  7. Gui F, You Z, Fu S, Wu H, Zhang Y. Endothelial Dysfunction in Diabetic Retinopathy. *Front Endocrinol (Lausanne)*. 2020 Sep 4;11:591. doi: 10.3389/fendo.2020.00591.
  8. Tang L, Xu GT, Zhang JF. Inflammation in diabetic retinopathy: possible roles in pathogenesis and potential implications for therapy. *Neural Regen Res*. 2023 May;18(5):976-982. doi: 10.4103/1673-5374.355743.
  9. Seo H, Park SJ, Song M. Diabetic Retinopathy (DR): Mechanisms, Current Therapies, and Emerging Strategies. *Cells*. 2025 Mar 4;14(5):376. doi: 10.3390/cells14050376.
  10. Wang J, Song X, Xia Z, Feng S, Zhang H, Xu C, et al. Serum biomarkers for predicting microvascular complications of diabetes mellitus. *Expert Rev Mol Diagn*. 2024 Aug;24(8):703-713. doi: 10.1080/14737159.2024.2391021.
  11. Szeto SK, Lai TY, Vujosevic S, Sun JK, Sadda SR, Tan G, et al. Optical coherence tomography in the management of diabetic macular oedema. *Prog Retin Eye Res*. 2024 Jan;98:101220. doi: 10.1016/j.preteyeres.2023.101220. Epub 2023 Nov 7. Erratum in: *Prog Retin Eye Res*. 2025 Jan;104:101319. doi: 10.1016/j.preteyeres.2024.101319.
  12. Nanji K, Hatamnejad A, Grad J, El-Sayes A, Mihalache A, Gemae M, et al. Visual outcomes associated with optical coherence tomography biomarkers in diabetic macular edema: A systematic review. *Surv Ophthalmol*. 2026 Mar-Apr;71(2):289-308. doi: 10.1016/j.survophthal.2025.09.009.
  13. Cappenberg A, Kardell M, Zarbock A. Selectin-Mediated Signaling-Shedding Light on the Regulation of Integrin Activity in Neutrophils. *Cells*. 2022 Apr 12;11(8):1310. doi: 10.3390/cells11081310. PMID: 35455989; PMCID: PMC9025114.
  14. Przędzek K, Cibor D, Zwolińska-Wcisło M, Owczarek D. Circulating cell adhesion molecules as biomarkers in inflammatory bowel disease: a systematic review and meta-analysis. *Front Immunol*. 2025 Dec 1;16:1680317. doi: 10.3389/fimmu.2025.1680317.
  15. Machoń NJ, Zdanowska N, Klimek-Trojan P, Owczarczyk-Saczonek A. Vascular Cell Adhesion Molecule 1 and E-Selectin as Potential Cardiovascular Risk Biomarkers in Psoriasis. *Int J Mol Sci*. 2025 Jan 18;26(2):792. doi: 10.3390/ijms26020792.
  16. Ekelund C, Dereke J, Nilsson C, Landin-Olsson M. Are soluble E-selectin, ICAM-1, and VCAM-1 potential predictors for the development of diabetic retinopathy in young adults, 15-34 years of age? A Swedish prospective cohort study. *PLoS One*. 2024 Jun 6;19(6):e0304173. doi: 10.1371/journal.pone.0304173.
  17. Kuo CYJ, Murphy R, Rupenthal ID, Mugisho OO. Correlation between the progression of diabetic retinopathy and inflammasome biomarkers in vitreous and serum - a systematic review. *BMC Ophthalmol*. 2022 May 27;22(1):238. doi: 10.1186/s12886-022-02439-2.
  18. Gericke A, Buonfiglio F. Physiological and Pathophysiological Relevance of Nitric Oxide Synthases (NOS) in Retinal Blood Vessels. *Front Biosci (Landmark Ed)*. 2024 May 16;29(5):190. doi: 10.31083/j.fbl2905190.
  19. Adly AAM, Ismail EA, Tawfik LM, Ebeid FSE, Hassan AAS. Endothelial monocyte activating polypeptide II in children and adolescents with type 1 diabetes mellitus: Relation to micro-vascular complications. *Cytokine*. 2015 Dec;76(2):156-162. doi: 10.1016/j.cyto.2015.06.006.
  20. Wilkinson CP, Ferris FL 3rd, Klein RE, Lee PP, Agardh CD, Davis M, et al.; Global Diabetic Retinopathy Project Group. Proposed international clinical diabetic retinopathy and diabetic macular edema disease severity scales. *Ophthalmology*. 2003 Sep;110(9):1677-82. doi: 10.1016/S0161-6420(03)00475-5.
  21. Yang Z, Tan TE, Shao Y, Wong TY, Li X. Classification of diabetic retinopathy: Past, present and future. *Front Endocrinol (Lausanne)*. 2022 Dec 16;13:1079217. doi: 10.3389/fendo.2022.1079217.
  22. Harding S, Greenwood R, Aldington S, Gibson J, Owens D, Taylor R, et al.; Diabetic Retinopathy Grading and Disease Management Working Party. Grading and disease management in national screening for diabetic retinopathy in England and Wales. *Diabet Med*. 2003 Dec;20(12):965-71. doi: 10.1111/j.1464-5491.2003.01077.x.
  23. Wilkinson CP, Ferris FL 3rd, Klein RE, Lee PP, Agardh CD, Davis M, et al.; Global Diabetic Retinopathy Project Group. Proposed international clinical diabetic retinopathy and diabetic macular edema disease severity scales. *Ophthalmology*. 2003 Sep;110(9):1677-82. doi: 10.1016/S0161-6420(03)00475-5.
  24. Bennett ST, Lehman CM, Rodgers GM, authors; Thompson C, Blaylock RC, editors. *Laboratory Hemostasis: A Practical Guide for Pathologists*. 2nd ed. Cham: Springer; 2015. 244 p. doi: 10.1007/0-387-36840-X.
  25. Miranda KM, Espey MG, Wink DA. A rapid, simple spectrophotometric method for simultaneous detection of nitrate and nitrite. *Nitric Oxide*. 2001 Feb;5(1):62-71. doi: 10.1006/niox.2000.0319.
  26. Kanda Y. Investigation of the freely available easy-to-use software 'EZR' for medical statistics. *Bone Marrow Transplant*. 2013 Mar;48(3):452-8. doi: 10.1038/bmt.2012.244.
  27. Hartnett ME, Fickweiler W, Adamis AP, Brownlee M, Das A, Duh EJ, et al. Rationale of Basic and Cellular Mechanisms Considered in Updating the Staging System for Diabetic Retinal Disease. *Ophthalmol Sci*. 2024 Mar 27;4(5):100521. doi: 10.1016/j.xops.2024.100521.
  28. Storti F, Pulley J, Kuner P, Abt M, Luhmann UFO. Circulating Biomarkers of Inflammation and Endothelial Activation in Diabetic Retinopathy. *Transl Vis Sci Technol*. 2021 Oct 4;10(12):8. doi: 10.1167/tvst.10.12.8.
  29. Yanxia C, Xiongyi Y, Min F, Xiaoyun K. Optical Coherence Tomography-Based Grading of Diabetic Macular Edema Is Associated with Systemic Inflammatory Indices and Imaging Biomarkers. *Ophthalmic Res*. 2024;67(1):96-106. doi: 10.1159/000535199.

30. Sen S, Khalid H, Udaya P, Raman R, Rajendram R, ElHousseini Z, et al. Ultrastructural imaging biomarkers in diabetic macular edema: A major review. *Indian J Ophthalmol*. 2025 Jan 1;73(Suppl 1):S7-S23. doi: 10.4103/IJO.IJO\_878\_24. Epub 2024 Dec 24. Erratum in: *Indian J Ophthalmol*. 2025 Feb 1;73(2):309. doi: 10.4103/IJO.IJO\_107\_25.
31. Serdyuk AV, Mogilevsky SYu, Babenko MS, Ziablitsev SV. Atherogenic dyslipidemia (lipid fractions and simple lipid indices) as a predictor of diabetic macular edema and adverse optical coherence tomography outcomes in diabetic retinopathy. *Mižnarodnij endokrinologičnij žurnal*. 2026;22(2):126-134. doi: 10.22141/2224-0721.22.2.2026.1686.
32. Siddiqui K, George TP, Mujammami M, Isnani A, Alfadda AA. The association of cell adhesion molecules and selectins (VCAM-1, ICAM-1, E-selectin, L-selectin, and P-selectin) with microvascular complications in patients with type 2 diabetes: A follow-up study. *Front Endocrinol (Lausanne)*. 2023 Feb 9;14:1072288. doi: 10.3389/fendo.2023.1072288.
33. Yang J, Liu Z. Mechanistic Pathogenesis of Endothelial Dysfunction in Diabetic Nephropathy and Retinopathy. *Front Endocrinol (Lausanne)*. 2022 May 25;13:816400. doi: 10.3389/fendo.2022.816400.
34. Sheng X, Zhang C, Zhao J, Xu J, Zhang P, Ding Q, et al. Microvascular destabilization and intricat network of the cytokines in diabetic retinopathy: from the perspective of cellular and molecular components. *Cell Biosci*. 2024 Jun 27;14(1):85. doi: 10.1186/s13578-024-01269-7.
35. Blum A, Pastukh N, Socca D, Jabaly H. Levels of adhesion molecules in peripheral blood correlat with stages of diabetic retinopathy and may serve as bio markers for microvascular complications. *Cytokine*. 2018 Jun;106:76-79. doi: 10.1016/j.cyto.2017.10.014.
36. Mogilevskii SI, Serdiuk AV, Zyablitsev SV. Prognostic biomarkers of non-proliferative diabetic retinopathy progression in type 2 diabetes mellitus. *Ukrainian Journal of Ophthalmology*. 2024;4(519):38-45. doi: 10.31288/oftalmolzh202443845.
37. Majidova SR. Evaluation of Hypoxia and Microcirculation Factors in the Progression of Diabetic Retinopathy. *Invest Ophthalmol Vis Sci*. 2024 Jan 2;65(1):35. doi: 10.1167/iovs.65.1.35.
38. Rajab HA, Baker NL, Hunt KJ, Klein R, Cleary PA, Lachin J, et al.; DCCT/EDIC Group of Investigators. The predictive role of markers of Inflammation and endothelial dysfunction on the course of diabetic retinopathy in type 1 diabetes. *J Diabetes Complications*. 2015 Jan-Feb;29(1):108-14. doi: 10.1016/j.jdiacomp.2014.08.004.
39. Muni RH, Kohly RP, Lee EQ, Manson JE, Semba RD, Schumberg DA. Prospective study of inflammatory biomarkers and risk of diabetic retinopathy in the diabetes control and complications trial. *JAMA Ophthalmol*. 2013 Apr;131(4):514-21. doi: 10.1001/jamaophthalmol.2013.2299.
40. Zheng X, Wan J, Tan G. The mechanisms of NLRP3 inflammasome/pyroptosis activation and their role in diabetic retinopathy. *Front Immunol*. 2023 Apr 25;14:1151185. doi: 10.3389/fimmu.2023.1151185.
41. van Horssen R, Eggermont AM, ten Hagen TL. Endothelial monocyte-activating polypeptide-II and its functions in (patho)physiological processes. *Cytokine Growth Factor Rev*. 2006 Oct;17(5):339-48. doi: 10.1016/j.cytogfr.2006.08.001.
42. Yuan X, Wang X, Ma X, Mao Y, Wang Q. AIMP1: multifunctional regulator in physiology and pathology with therapeutic implications. *PeerJ*. 2025 Nov 18;13:e20334. doi: 10.7717/peerj.20334.
43. Mogilevskyy S Yu, Serdiuk AV, Serdiuk VN, Ziablitsev SV. Model for predicting the efficacy of treating diabetic retinopathy in type 2 diabetes on the basis of determination of markers of endothelial dysfunction. *Journal of Ophthalmology (Ukraine)*. 2025;5(527):3-12. doi: 10.31288/oftalmolzh20256312.
44. Monickaraj F, Acosta G, Cabrera AP, Das A. Transcriptomic Profiling Reveals Chemokine CXCL1 as a Mediator for Neutrophil Recruitment Associated With Blood-Retinal Barrier Alteration in Diabetic Retinopathy. *Diabetes*. 2023 Jun 1;72(6):781-794. doi: 10.2337/db22-0619.
45. Guo C, Sodhi A. Molecular stress and neurovascular injury in the diabetic retina. *J Clin Invest*. 2026 Mar 2;136(5):e200945. doi: 10.1172/JCI200945.
46. Kaštelan S, Orešković I, Bišćan F, Kaštelan H, Gverović Antunica A. Inflammatory and angiogenic biomarkers in diabetic retinopathy. *Biochem Med (Zagreb)*. 2020 Oct 15;30(3):030502. doi: 10.11613/BM.2020.030502.